

Caudals and Calyces: The Curious Case of a Consumed Chiapan Colubroid

Author(s): Jonathan A. Campbell, Eric N. Smith, and Alexander S. Hall Source: Journal of Herpetology, 52(4):459-472. Published By: The Society for the Study of Amphibians and Reptiles <u>https://doi.org/10.1670/18-042</u> URL: http://www.bioone.org/doi/full/10.1670/18-042

BioOne (<u>www.bioone.org</u>) is a nonprofit, online aggregation of core research in the biological, ecological, and environmental sciences. BioOne provides a sustainable online platform for over 170 journals and books published by nonprofit societies, associations, museums, institutions, and presses.

Your use of this PDF, the BioOne Web site, and all posted and associated content indicates your acceptance of BioOne's Terms of Use, available at www.bioone.org/page/terms_of_use.

Usage of BioOne content is strictly limited to personal, educational, and non-commercial use. Commercial inquiries or rights and permissions requests should be directed to the individual publisher as copyright holder.

BioOne sees sustainable scholarly publishing as an inherently collaborative enterprise connecting authors, nonprofit publishers, academic institutions, research libraries, and research funders in the common goal of maximizing access to critical research.

Caudals and Calyces: The Curious Case of a Consumed Chiapan Colubroid

JONATHAN A. CAMPBELL,^{1,2} ERIC N. SMITH,¹ AND ALEXANDER S. HALL^{1,3}

¹Department of Biology, University of Texas at Arlington, Arlington, Texas USA

ABSTRACT.—A new genus and species of colubroid snake is described from the isolated highlands of western Chiapas. This enigmatic little snake possesses a unique suite of characters that defies placing it in any known genus and clearly distinguishes it from all known genera. Several of the most unusual features include subcaudals undivided throughout the length of the tail and a simple hemipenis completely adorned with calyces and having a sulcus spermaticus that remains unbifurcated until the apical portion of the organ. Neither of these characteristics is known for any other colubroid of the Western Hemisphere. Consideration of morphology places the new snake in the Dipsadidae and suggests that *Adelphicos, Atractus, Geophis,* and *Chapinophis* are among its closest relatives.

RESUMEN.—Se describe un nuevo género y especie de serpiente colubrina de las montañas aisladas del oeste de Chiapas. Esta enigmática y pequeña serpiente posee un conjunto único de caracteres que impide su asignación a cualquier otro género y claramente la distingue de cualquier género conocido. Varios de las características inusuales incluyen subcaudales no divididas a lo largo de toda la cola y un hemipene simple, completamente cubierto de cálices, y con un surco espermático que permanece no bifurcado hasta el ápice del órgano. Ninguna de estas características es conocida de cualquier colúbrido en el hemisferio occidental. Examen de morfología sugiere que la nueva serpiente pertenece a la familia Dipsadidae y su afinidad esta con *Adelphicos, Atractus, Geophis, y Chapinophis*.

Prey items, especially small snakes, are frequently encountered in stomachs of Coralsnakes (Schmidt, 1932). Indeed, in several instances, new species of snakes have been recovered from Coralsnakes of the genus *Micrurus. Geophis dunni* Schmidt (1932) was removed from an individual of *Micrurus nigrocinctus* from Nicaragua, and this species has remained an enigma. The holotype was collected during or before 1909 (Townsend, 2006) from a region of Nicaragua that is thought to have become relatively well surveyed. Nevertheless, *G. dunni* has remained known from only the single specimen. In another instance of predation leading to discovery of a taxonomic novelty, Hay (1892) described *Storeria victa* (*Storeria dekayi victa*) that he encountered in the stomach of a *Micrurus fulvius* from Florida.

Coralsnakes are undoubtedly far more efficient predators of certain species of small snakes than human collectors, as revealed by a cursory survey of secretive or fossorial species of Middle American snakes found in their stomachs. Numerous species of colubroids are recorded from no more than a handful of individuals, and some are known from only their original descriptions: *Adelphicos daryi* Campbell and Ford (1982), *Adelphicos ibarraorum* Campbell and Brodie (1988), *Chapinophis xanthocheilus* Campbell and Smith (1998), *Geophis isthmicus* Downs (1967), *Geophis juarezi* Nieto-Montes de Oca (2003), *Geophis maculiferus* Downs (1967), and *Rhadinophanes monticola* Myers and Campbell (1981) are but a few examples of rare species becoming known in recent decades.

We were not surprised when a small snake was recovered from the stomach of a *M. nigrocinctus* from southern Mexico. Following the key to Mexican genera of snakes in Smith and Taylor (1945), this snake is identified as *Geophis*, which—on the basis of a suite of characters—it is not. Subsequent examination clearly indicated that this prey item represented a previously unknown snake, unlike anything currently recognized from the New World tropics. This small snake was obtained now over 40 yr ago, and the report of its discovery has been a long time in coming. We were optimistic that additional specimens might be secured, but after at least a dozen more trips into the region spanning several decades, we have been unrewarded.

The colubroids of the Western Hemisphere currently are allocated into three families: Natricidae, Colubridae, and Dipsadidae (Vidal et al., 2007, 2010; Zaher et al., 2009; sensu Grazziotin et al., 2012), although these families are sometimes placed at the subfamily level (Figueroa et al., 2016). Regardless of taxonomic rank, these groups of snakes are recognizable by distinctive molecular differences. Morphological characteristics that clearly define individual groups are more difficult to categorize owing to the large number of species and morphologically diverse assemblages. The largest American clade of snakes, the Dipsadidae, is composed of almost 100 genera in over 700 species that have radiated into practically every terrestrial and freshwater habitat.

Several clades within the Dipsadidae are currently placed at the subfamily level (Vidal et al., 2007, 2010; Zaher et al., 2009). The Xenodontinae contains genera that are mostly South American in distribution, possess enlarged spines on the sides of a bilobed hemipenial body, and the lobes bear two distinctly ornamented regions: a sulcate surface bearing a capitulum and the asulcate surface with enlarged spinulate or papillate calyces, or sometimes nude; further, the sulcus spermaticus bifurcates near the base of the organ (Zaher, 1999). The Dipsadidae is primarily a Middle American lineage containing over 25 genera lacking enlarged lateral spines on the hemipenial body and do not have two different ornamented regions on the lobes. Further, the organ may be single or bilobed and may be noncapitate or semicapitate. Most species have a capitate, calyculate hemipenis with a sulcus spermaticus bifurcating within the capitulum. A synapomorphy supporting this group is a sulcus spermaticus bifurcating at the base of, or within, the capitulum (Cadle, 1984c; Zaher, 1999; Myers and Cadle, 2003). The hemipenis of the new genus described here is unlike that of any known colubroid.

MATERIALS AND METHODS

Protocols for making most standard scale counts follow Myers (1974). We determined the number of ventral scutes

²Corresponding Author. E-mail: campbell@uta.edu

³Present address: Thermo Fisher Scientific, 16700 Park Row Drive, Houston, Texas USA

DOI: 10.1670/18-042

using the procedure suggested by Dowling (1951), in which the anteriormost ventral is that bordered on either side by the first row of dorsal scales. In many snakes there is a single scale between the nasal(s) and eye. A single scale, when present in dipsadids, is longer than high and therefore referred to as a loreal (Peters, 1964). We measured the body and tail with a standard meterstick to the nearest 1 mm; smaller structures such as the head and hemipenis were measured to the nearest 0.1 mm using a Vernier caliper. We obtained locations and distances using topographic maps (1:250,000) issued by the Instituto Nacional de Estadística, Geografía e Informática (2000) and Google Earth Pro (2017). Specimens were preserved by injection with formalin diluted to 10% of stock solution and then transferred to 70% ethanol for permanent storage. Fortunately, enough formalin was injected into the body cavity to stop the digestive process of the Coralsnake that preved on the snake (the primary subject of this paper).

We prepared the hemipenis of the new taxon and comparative material using the technique of Pesantes (1994), with modifications as suggested by Myers and Cadle (2003) and Zaher and Prudente (2003), expanding everted hemipenes by introducing a syringe needle with a blunt tip at the base, tying the organ at this level, and injecting heated petroleum jelly with blue wax-dye through the needle until maximal possible expansion was achieved without risking rupture.

The artwork reconstructing the new taxon as it probably appeared in life was accomplished by Gabriel Ugueto, who used a series of photographs and measurements of the holotype as a reference. The illustration was drawn and painted digitally in Corel Painter and Adobe Photoshop using a Wacom Cintiq Pen Display.

To compare cranial osteology of small, rare, and delicate specimens we collected computed tomography (CT) data from skulls of the new taxon and comparative material. Formalinfixed specimens were immobilized in a combination of floral foam, plastic straws or cups, and tape. Then, each specimen (sometimes two to four specimens simultaneously) were CT scanned at the University of Texas Arlington Shimadzu Center for Environmental Forensics and Material Science using a Shimadzu inspeXio SMX-100CT (Shimadzu, Inc., Kyoto, Japan). We collected CT data using several parameter combinations deemed suitable to preserved snake specimens. Each scan lasted about 15–30 min.

We used Shimadzu's inspeXio software to reconstruct raw Xray data and exported them as stacks of $1,024 \times 1,024$ 16-bit TIFF images. Each stack was rotated and cropped in ImageJ and imported into AvizoTM Software v.8.1 and 9.4 (Thermo Fisher Scientific, Inc., Waltham, Massachusetts USA). We isolated and converted skulls into one surface per skull using interactive thresholding. These surface files were the basis for collecting characteristics across the new taxon and comparative taxa. For the new taxon and *Adelphicos quadrivirigatus* (UTA R-22680), we manually segmented the lower jaw bones and the maxilla, ectopterygoid, pterygoid, and quadrate. Because of the digested state of the new taxon, reasonable segmentation of these bones was possible on only one side of the skull.

We examined comparative material from the Amphibian and Reptile Diversity Research Center at University of Texas Arlington, which contains representatives of almost every genus of Middle American snakes. For osteological comparisons, we prepared CT scans for the following species of dipsadids and colubrids (all numbers refer to UTA, except for UF H-157664 and AMNH 103068): *Adelphicos latifaciatus* (R- 12245–12246), A. quadrivirgatus (R-22680), Amastridium veliferum (14146), Atractus lancini (R-59423), Atractus. univittatus (R-3600), Atractus trilineatus (R-22264), C. xanthocheilus (R-37591), Chersodromus leibmanni R-14132, 52625), Coniophanes fissidens (R-9413), Coniophanes imperialis (R-4328), Conophis vittatus (R-52631), Cryophis hallbergi (R-12272, 52632), Diadophis punctatus (R-5952), Dipsas indica (R-18175), Enuliophis sclateri (R-45181), Enulius flavitorques (R-56376), Ficimia publia (R-63119), Geagras redimitus (R-26690), Geophis nigrocinctus (R-5908), Geophis omiltemanus (R-5055), Hydromorphus concolor (R-44940, R-44941), Hypsiglena torquata (R-57106), Imantodes cenchoa (R-39231), Leptodeira annulata (R-59214), Leptodeira septentrionalis (R-14487), Leptodeira uribei (R-57688), Manolepis putnami (R-53007), Ninia diademata (R-12371), Ninia pavimentata (R-42407), Nothopsis rugosus, (R-40098), Oxybelis aeneus (R-53369), Omoadiphas aurula (UF H-157664), Pliocercus elapoides (R-52569), Pseudoleptodeira latifasciata (R-59558), Rhadinaea fulvivittis (R-4692), Rhadinaea taeniata (R-4424), Rhadinella hannsteini (R-21720), Rhadinophanes monticola (R-63277), Rhinocheilus lecontei (R-50696, 57755), Sibynomorphus mikanii (R-37712), Sibon carri (R-37270), Sibon nebulata (R-4700), Tantalophis discolor (ANMH-103068), Tantilla rubra (R-12455), Tantilla shawi (R-36810), Tretanorrhinus nigroluteus (R-52336), Trimetopon pliolepis (R-44969), Tropidodipsas fasciata (R-52645), and Tropidodipsas philippii (R-57499). To view the entire specimen, we collected radiographs from the new taxon using the SMX-100CT and exported them as $1,024 \times 1,024$ 16-bit grayscale TIFF files. Each radiograph ran at 35 kV and 40 μA and averaged over 999 integrations in inspeXio. Original tomograms and surface files for the new taxon and Adelphicos quadrivirgatus (R-22680) are available on Morphosource (http://www.morphosource.org/ Detail/ProjectDetail/Show/project_id/555) and archived at the University of Texas at Arlington Amphibian and Reptile Diversity Research Center digital image collection.

The following sources have also been helpful in comparing the new genus with other Middle American snakes with regard to scalation, osteology, internal morphology, and biogeography: Hay (1892), Cope (1894), Smith (1941), Tanner (1943, 1944), Burger and Werler (1954), Duellman (1958a,b), Dowling and Savage (1960), Leviton and Tanner (1960), Peters (1960), Bogert and Duellman (1963), Dowling (1967), Downs (1967), Wilson and Meyer (1969), Myers (1974, 1982, 1984, 2003), Cundall (1981), Myers and Campbell (1981), Campbell and Ford (1982), Cadle (1984a,b,c), Jenner and Dowling (1985), Kofron (1985a,b, 1987), Campbell and Brodie (1988), Campbell (1989), Crother (1989), Smith and Campbell (1996), Campbell and Smith (1998), Zaher (1999), Smith and Chiszar (2001), Savage (2002), Mulcahy (2003, 2007), Myers and Cadle (2003), Nieto-Montes de Oca (2003), Schargel and Castoe (2003), Pinou et al. (2004), Townsend (2006), Cundall and Irish (2008), Harvey et al. (2008), McDowell (2008), Passos et al. (2009, 2013), Mulcahy et al. (2011)Angarita-Sierra (2014), Pietro et al. (2014), Zaher et al. (2014), Klaczko et al. (2016), and Canseco-Márquez et al. (2018).

Systematic Account

We are aware of no single morphological character that clearly differentiates the Dipsadidae from the Colubridae. We are confident, however, in placing the new taxon described below in the Dipsadidae on the basis of overall gestalt and similarity of certain characters with Middle American genera (*Adelphicos, Atractus, Chapinophis,* and *Geophis*) such as elongated braincase, loreal entering the orbit, prefrontal making contact



FIG. 1. Cenaspis aenigma (holotype, UTA R-10544, 258 mm total length), drawing reconstructing species in life.

with the eye, and maxilla with relatively few teeth (14–15) and probably lacking enlarged "rear-fangs."

Cenaspis, new genus (Figs. 1–7)

Type Species.—Cenaspis aenigma, new species (described below). Diagnosis of Genus.-This genus differs from all other New World dipsadids by two unique features: 1) all subcaudals undivided and 2) hemipenis single, noncapitate, hemipenial body and apical region completely covered with calyces, and sulcus spermaticus simple. Rhinocheilus is the only other American colubroid north of Panama having undivided subcaudals, distinguished from Cenaspis in always having at least a few divided subcaudals, preocular present, dorsals in 23 rows at mid-body and reduced to 19 posteriorly, and a reddish and black pattern with dorsal blotches or bands. Although some other New World genera have single hemipenes, in every case of which we are aware, the hemipenial body bears spines or spinules (vs. calyces), there usually is distinct capitation, and the sulcus spermaticus is bifurcate for much of the length of the capitulum (Tables 1, 2).

Etymology.—The generic name is derived from the Latin *cena*, meaning dinner, and *aspis*, meaning a kind of snake, in reference to predation on the single known individual of this snake. The name taken literally means "dinner snake."

Cenaspis aenigma, new species

Holotype.—UTA R-10544, an adult male, obtained on "La Loma," located some 20–25 km (by road) W-NW of Rizo de Oro (sometimes known as Nueva Tenochtitlán), Chiapas, Mexico. Access is gained into this region by means of a logging road connecting Rizo de Oro with Colonia Rodulfo Figueroa; the latter is a small settlement very near the border with Oaxaca. The type locality lies to the N-NW of Colonia Rodulfo Figueroa on the western slopes of Cerro El Baúl, the highest peak in the region, rising to an elevation of 2,050 m. Collected by the late Julio Ornelas-Martínez on 6 July 1976.



FIG. 2. *Cenaspis aenigma* (holotype, UTA R-10544, 258 mm total length), dorsal (A) and ventral (B) aspects of preserved specimen.



FIG. 3. *Cenaspis aenigma* (holotype, UTA R-10544), dorsal (A), lateral (B), and ventral (C) aspects of head of preserved specimen.

Diagnosis.—Differs from other Middle American genera of dipsadids except *Adelphicos, Atractus, Chapinophis, Chersodromus, Geophis, Ninia,* some *Sibon* in having prefrontal enter orbit. *Chersodromus* and *Ninia* (characters for *Cenaspis aenigma* in parentheses) have keeled (vs. smooth) dorsal scales; *Adelphicos* and *Sibon* have ≤ 15 (vs. 17) dorsal scale rows at mid-body; *Atractus, Chersodromus, Geophis,* and *Sibon* have 1 + 2 or 0 + 1 (vs. 1 + 1) temporals; *Adelphicos, Chapinophis,* and *Geophis* usually have ≤ 7 (vs. 8) supralabials; and *Adelphicos, Chapinophis,* and *Trimetopon* have a divided cloacal scute (vs. undivided) (Tables 1, 2). For additional characteristics see diagnosis for genus.

Description of the Holotype.--Moderately slender, small snake with snout-vent length 222 mm, tail length 36 mm, and total length 258 mm (Figs. 1, 2); head length 9.0 mm, eye diameter 1.0 mm, snout-to-eye distance 1.2 mm (Fig. 3); portions of head on left side, ventral surface, and neck digested and scales missing; internasals small, less than half size of prefrontals; prefrontals relatively large, anterolateral portion forming upper anterior border of eye; frontal 2.4 long \times 1.8 wide, flanked by large supraoculars; parietals long, tapering, and becoming slender posteriorly; nasal scale single apparently without vertical suture; single horizontally elongate loreal extending posteriorly to border anterior of orbit (alternatively, this scale might be considered a preocular with loreal missing, but see Peters, 1964); supralabials 8/8, supralabials 4–5 contacting lower border of orbit; postoculars 2/2, upper long and narrow, about twice as large as the lower; temporals 1 + 1 + 2; dorsal scale rows at midbody 17, unreduced posteriorly; ventrals ~140, some missing anteriorly on body but estimated on the basis of segmental (vertebral) number; cloacal scute undivided; subcaudals 36,



FIG. 4. *Cenaspis aenigma* (holotype, UTA R-10544), volume rendering of dorsal (A), lateral (B), and ventral (C) aspects of skull.

undivided; tail spine sharply pointed, about as long as preceding 2 subcaudals; mid-length of tail encircled by 8 scales between subcaudals.

Dorsum without conspicuous pattern; dorsal ground color medium brown, grading to paler brown on sides; anterior supralabials to level of eye white, posterior lower portion of supralabials white; apical portion of most dorsal scales with dark brown spot, especially on middle half of body; cranial end of individual dorsal scales also dark brown over much of body (Figs. 1, 2A); venter of body cream with three series of dark brown triangular blotches on ventrals creating irregular striping; tail cream with single series of mid-ventral blotches (Fig. 2B).

Premaxilla mostly absent (Figs. 4, 5A–C), perhaps owing to digestion, but prominent ascending process oriented vertically and attaching dorsally to septomaxilla and nasals; nasals in broad medial contact, longer than wide with broad downwardly curving lateral winglike processes, rounded posteriorly, in contact with frontals; prokinetic joint attaching snout with braincase formed by articulation of nasals and septomaxillae with frontals; prefontals robust, blocklike, anterior edge concave without anteriorly projecting processes, ventrally long articulating surface with maxillae, extended by posteroventral



FIG. 5. Comparison between segmented skulls of *Cenaspis aenigma* (UTA R-10544) (A–C) and *Adelphicos quadrivirigatus sargii* (UTA R-22680) (D–F). Dorsal (A and D), lateral (B and E), and ventral (C and F) aspects; quadrate shown in red, palatomaxillary arch in yellow, mandible in green, and cranium in blue. The quadrate of the *C. aenigma* holotype was digested on the left side, so the right one is shown (although somewhat digested).

process; paired frontals with broad median suture, posteriorly broad articulation with parietal forming U shape; parietals united, extending downward to basisphenoid; supraoccipitals relatively large, lacking transverse crest, with median keel; postorbital large, well-developed, articulating broadly with parietal, reaching frontal, bordered dorsally by distinct parietal ridge, extending ventrally to occupy most of posterior margin of orbit, nearly reaching ectopterygoid; dentigerous bones of the palatomaxillary arch include maxillary, palatine, and pterygoid (Fig. 6A); in lower jaw dentary also with teeth (Fig. 6B); teeth short, conical, blunt, only slightly recurved; maxilla long, almost straight, extending to level about equal to posterior edge of eye, broad palatine process projecting slightly posteromedially and medial edge with blunt point; ectopterygoid process slightly more narrow than palatine process, angling posteriorly from terminus of bone, medial edge straight; number of maxillary teeth 14/14 (anterior tip of maxillae broken, possibly accounting for one additional tooth on each side); increase in tooth socket size from anterior to posterior indicating corresponding increase in tooth size, no posterior diastema; anterior end of ectopterygoid relatively short, strongly bifurcating with lateral fork about twice as long as medial fork, extending posteriorly to level about equal to lateral flexure of pterygoid; palatine teeth

5/4, tooth sockets large; pterygoid teeth 16/17, decreasing in size posteriorly; supratemporal (right only, left missing) relatively short, straight, and wide, extending just barely beyond caudal edge of braincase, extending anteriorly only to prootic; quadrate short, stout, roughly triangular, well-developed heterocoelous condyle articulating with mandible, dorsally flattened and flared dorsoanteriorly, large oval foramen in upper posterior of bone (Fig. 6C).

The right hemipenis in situ extends to subcaudal 12; left hemipenis removed, everted, and expanded, total length from base to apex ~10 mm; organ mostly cylindrical, distally becoming very slightly bulbous; sulcus spermaticus single through most of length, becoming bifurcated at apex (level of subcaudal 12); except for extreme lower base, entire organ calyculate, with large calyces proximally (alternately being considered flounced because of relatively few vertical crossridges), becoming smaller distally; as few as 3–4 calyces encircling organ near base, 18–19 near apex; micro-ornamentation of ridges papillate; apex not differentiated (Fig. 7).

Etymology.—From the Latin *aenigma* meaning a riddle or mystery.

Comparison with Other Middle American Taxa.—About half of the genera of Dipsadidae have a preocular contacting the supra-



FIG. 6. Comparison between segmented bones of *Cenaspis aenigma* (UTA R-10544) (A–C) and *Adelphicos quadrivirigatus sargii* (UTA R-22680) (D–F). Palatomaxillary arch (A and D; yellow), mandible (B and E; green), and quadrate (C and F; red). Left palatomaxillary arches shown in dorsal, lateral, medial, and, ventral views (top to bottom); left mandibles shown in dorsal, medial, lateral, and ventral views (top to bottom); right quadrate shown for *C. aenigma* (C) in lateral (left) and medial (right) views, and left quadrate for *A. quadrivirigatus sargii* (F) in medial (left) and lateral (right) views.

ocular, precluding contact of the prefrontal with the orbit (Table 1). When the preocular is absent as in *Cenaspis, Adelphicos, Atractus, Chapinophis, Chersodromus, Enuliophis, Enulius, Geophis, Ninia, Omoadipsas,* and some *Sibon,* the prefrontal enters the eye. Associated with this trait is a single, horizontally elongate loreal

between the nasal(s) and orbit. Most often small burrowing or terrestrial Dipsadidae have 15 to 17 rows of smooth dorsal scales at mid-body and these are unreduced posteriorly; exceptions are *Chersodromus* and *Ninia*, which have keeled scales. Genera with 19 or more scale rows at mid-body (*Coniophanes, Cryophis*,

NEW COLUBRID GENUS AND SPECIES FROM CHIAPAS



FIG. 7. Selected hemipenes of Nuclear Central American snakes. (A) *Adelphicos latifasciatus* from Oaxaca, UTA R-12246 (left hemipenis, 204 mm snout–vent length [SVL], 54 mm tail length); (B) *Cenaspis aenigma* holotype from Chiapas, UTA R-10544 (left hemipenis, 222 mm SVL, 36 mm tail length); (C) *Cryophis hallbergi* from Oaxaca, UTA R-12272 (left hemipenis, 478 mm SVL, 137 mm tail length); (D) *Geophis rhodogaster* from Guatemala, UTA R-28349 (right hemipenis, 223 mm SVL, 59 mm tail length). Sulcate (left) and asulcate (right) views shown. Note that, except for the hemipenis of *C. aenigma*, all other organs are spinous and show a well differentiated capitulum, slightly bilobed in *Geophis rhodogaster*.

Eridaphas, Hypsiglena, Leptodeira, Pseudoleptodeira, Rhadinophanes, and *Tretanorhinus*) generally have scale-row reductions on the posterior of the body.

Few other Middle American snakes possess a ventral pattern resembling that of *Cenaspis aenigma*. The venter is immaculate in most species, although colors may grade into different hues from anterior to posterior of body, and often the dorsal coloration encroaches on the lateral edges of ventrals. *Coniophanes quinquevittatus* and *Coniophanes bipunctatus* have distinct dark round spots near the outer edge of each ventral and the midventer is unmarked. *Ninia diademata* frequently has three regular series of dark markings on the ventrals: a series is present on the lateral portion of each ventral and a large wellformed roundish to triangular spot midventrally. Unlike *Cenaspis aenigma*, in which the markings in all series are approximately the same size and shape, in *N. diademata* the midventral series is conspicuously larger and differently shaped from the lateral series (Burger and Werler, 1954). *Ninia maculata*

and *Ninia psephota* usually have dark ventral markings that may variably be in a single midventral series, two series near lateral edges of ventrals, or as an irregular checkerboard, but rarely dark markings are arranged in more-or-less three series (lateral and midventral), although in irregular lines (Smith and Campbell, 1996; Savage, 2002).

Many genera of the Dipsadidae have two enlarged teeth on the posterior of the maxilla that may be grooved and often set off from more anterior teeth by a diastema (e.g., *Amastridium*, *Coniophanes, Imantodes, Leptodeira, Tantalophis;* Table 2). In *Cenaspis* 14–15 maxillary teeth appear to decrease in size posteriorly (on the basis of tooth socket size) and no diastema or enlarged grooved teeth are present (Figs. 5, 6). Fewer maxillary teeth characterize *Adelphicos* (7–11), *Atractus* (5–11), *Chapinophis* (12), and *Chersodromus* (7–9), whereas more teeth are present in *Ninia* (15–18) and *Tretanorhinus* (27–30). Species of *Geophis* differ by having relatively longer, sharper, more slender teeth in contrast to the short, stout, and blunt teeth in *Cenaspis*. 466

TABLE 1. Comparison of certain scalation characters among Middle American dipsadid snakes. Some meristic characters subject to variation; usual condition given. Dark shade represents characters similar to *Cenaspis*; light shade represents characters at variance.

Taxon	Prefrontals	Preocular(s)	Supralabials	Temporals	Dorsal midbody scales, posterior condition	Dorsal scale ornamentation	Cloacal scute	Subcaudals
Cenaspis aenigma	Entering	Absent	8	1 + 1	17, upreduced	Smooth	Undivided	Undivided
A delphicos (veraepacis group)	Entering orbit	Absent	7	1 + 1	15, unreduced	Smooth	Divided	Divided
A delphicos (quadrivirgatus group)	Entering orbit	Absent	7	1 + 1	15, unreduced	Smooth	Divided	Divided
Amastridium	Not entering	Present	7	1 + 2	17, unreduced	Smooth	Divided	Divided
Atractus	Entering orbit	Absent	5–9	1 + 2	15–17, unreduced	Smooth	Undivided	Divided
Chapinophis	Entering	Absent	7	1 + 1	17, unreduced	Smooth	Divided	Divided
Chersodromus	Entering	Absent	7–8	1 + 2	15–17,	Keeled	Undivided	Divided
Coniophanes	Not entering orbit	Present	7	1 + 2	17–25, reduced	Smooth	Divided	Divided
Cryophis	Not entering	Present	9	2 + 2	23,	Keeled	Divided	Divided
Dipsas articulata	Variable	Variable ¹	6–11	Variable	13–19,	Smooth	Undivided	Divided
Enuliophis	Entering	Absent	6–7	1 + 1 1 + 2	15,	Smooth	Divided	Divided
Enulius	Entering orbit	Variable	5–8	1 + 2 1 + 1 1 + 2	15–17, reduced or	Smooth	Divided	Divided
Geophis (omiltemanus	Entering orbit	Absent	6 ²	1 + 2	15–17, unreduced	Smooth or keeled	Undivided	Divided
group) Geophis (sieboldi group)	Entering orbit	Absent	6 or fewer	0 + 1	15–17, unreduced	Keeled (at least above vent)	Undivided	Divided
Hydromorphus	Not entering	Variable ³	Usually 6	1 + 2	15–17, roducod	Smooth	Divided	Divided
Hypsiglena (including Eridinhas)	Not entering orbit	Present	7–8	1 + 2	19–23, reduced	Smooth	Divided	Divided
Imantodes	Not entering orbit	Present	8–9	1 + 2 2 + 2	17, reduced or	Smooth	Usually divided ⁴	Divided
Leptodeira	Not entering	Present	7–9, usually 8	1 + 2	17–25,	Smooth	Divided	Divided
Ninia	Entering	Absent	6–7	1 + 2	17–19,	Keeled	Usually	Divided
Nothopsis	Not entering	Present	9–13	Multiple	26–30,	Keeled	Undivided	Divided
Omoadiphas	Entering	Absent	6–7	0 + 1	17,	Smooth	Divided	Divided
Pseudoleptodeira	Not entering orbit	Present	8	1 + 2	23, reduced	Smooth	Divided	Divided
<i>Rhadinaea</i> (<i>decorata</i> group)	Not entering orbit	Present	8–9	1 + 2	17, unreduced	Smooth to weak keels	Divided	Divided

TABLE 1. Continued.

Taxon	Prefrontals	Preocular(s)	Supralabials	Temporals	Dorsal midbody scales, posterior condition	Dorsal scale ornamentation	Cloacal scute	Subcaudals
Rhadinaea	Not entering	Present	8	1 + 1	17,	Smooth	Divided	Divided
(<i>taeniata</i> group)	orbit			1 + 2	unreduced			
Rhadinella	Not entering orbit	Present	7–8	1 + 1 1 + 2	17–21, unreduced	Smooth	Divided	Divided
Rhadinophanes	Not entering orbit	Present	8	1 + 2	19, reduced	Smooth	Divided	Divided
<i>Sibon annulatus &</i> <i>S. argus</i> groups	Entering orbit ⁵	Absent	7–9	$1 + 2^{6}$	13–15, unreduced ⁷	Smooth	Undivided	Divided
Tantalophis	Not entering orbit	Present	7–8	1 + 2	19–21, reduced	Smooth	Divided	Divided
Tretanorhinus	Not entering orbit ⁸	Present	7–9	1 + 2	19–21, reduced	Keeled	Divided	Divided
Trimetopon	Not entering orbit	Present	7–8	1 + 1	15–17, unreduced	Smooth	Divided	Divided
Tropidodipsas	Not entering orbit	Present	6–7	1 + 2	17, unreduced	Weakly keeled	Undivided	Divided
<i>Urotheca</i> (now included within <i>Pliocercus</i>)	Not entering orbit	Present	6–8	1 + 1 1 + 2	17, unreduced	Smooth	Divided	Divided

¹ When present, preocular located above loreal, which enters eye.

² Supralabials 7 in the enigmatic *Geophis isthmicus*.

³ Preoculars 0–2; when absent loreal contacting orbit.

⁴ Cloacal plate usually divided in northern species; often entire in lower Central American and South American species.

⁵ In all species except *S. sanniolus*.

⁶ Temporals in *Sibon carri* 0 + 1.

⁷ Sibon anthracops has 13 dorsal scale rows at mid-body.
⁸ Prefontals often coalescing with loreal(s); number of loreals 0–2.

The ectopterygoid is slenderer in *Geophis* than in *Cenaspis* and usually less forked; only in some species in the *Geophis mutitorques* group does bifurcation of the anterior end with lateral anterior branch of the ectopterygoid approach the

condition seen in Cenaspis. Numerous other osteological features distinguish various species of dipsadids from C. aenigma. In Adelphicos latifasciatus, the prefrontal has two (middle and lower) distinctive forward processes, a more strongly defined dorsolateral parietal ridge, no foramen in the quadrate, and 10-11 short, stout maxillary teeth. Adelphicos quadrivirgata (Figs. 6D,E,F) is similar to A. latifasciatus except there is only a single middle forward prefrontal process (middle; Fig. 5E). In Amastridium veliferum the braincase is shorter; the postocular is narrow dorsally, becoming heavy at midlength, continuing to ventral end; and there are 13-14 + 2 maxillary teeth (enlarged, diastema present). Atractus lancini and A. univittatus have 11-14 long, sharp maxillary teeth, becoming smaller posteriorly; the prefrontal has an anteriorly projecting flange. There are 11 long, robust maxillary teeth, becoming smaller posteriorly in Chapinophis xanthocheilus; the postocular is stout, the prefrontal has a middle anterior projection, the anterior edge of frontal has median projections, and the prefrontal distinctly curves over the dorsum of skull. Chersodromus liebmanni has a relatively short braincase, a strong dorsolateral ridge is present on parietals, postoculars are absent, the prefrontal has three forward projections, and almost the anterior third of the maxilla is edentate, followed by about nine long needlelike teeth. Coniophanes imperialis has a short braincase, the prefontals have a large forward flange, the dorsolateral ridge on parietal is well developed, the supratemporal extends forward to almost the parietal, and the maxilla has 11-12 + 2 teeth that are posteriorly enlarged and grooved, separated by a diastema from more anterior teeth. Conophis vittatus has a short braincase; a postocular is present but not contacting frontal, the quadrate is relatively long and slender, the prefrontal has a forward-projecting flange, the maxilla bears 10 + 3 teeth with posterior teeth greatly enlarged and grooved, and a diastema is present. In Cryophis hallbergi the premaxilla does not reach the nasals, the prefrontals have a broad forward projecting flange, the postorbital does not reach the frontal, the supratabular extends forward to the parietal, the maxilla bears 21 + 2 teeth with posterior teeth enlarged and grooved, and a narrow diastema is present. Diadophis punctatus has a prefrontal with a middle forward-projecting process, the dorsolateral ridge of the parietal is well developed, the supratabular extends forward on to the parietal, the maxilla bears 8-9 + 2 teeth with posterior teeth enlarged, and a diastema is present. In Ficimia publia, the premaxilla is distinctly forward projecting, the prefrontal has single large anterior projection, and the maxilla bears 12-13 short stout teeth. In Geagras redimitus, (a small burrowing Mexican colubrid), the premaxillary is produced forward with a knoblike process that

TABLE 2. Comparison of selected morphological features among Middle American dipsadid snakes. Dark shade represents characters similar to Cenaspis; light shade represents at variance, unshaded represents comparative condition unknown in Cenaspis.

Taxon	Pupil shape	Maxilla teeth number ¹	Size maxillary teeth (very thin, thin, moderate, stout, very stout, extremely stout)	Maxillary diastema	Maxillary teeth grooved	Hemipenial shape	Truncus region ornamentation	Apical region ornamentation	Capitation	Sulcus spermaticus
Cenaspis aenigma	Subcircular	14–15	Unknown	No	Unknown	Single	Calyculate	Calyculate	Noncapitate	Simple, with
										fork
A delphicos	Round	7–11	Stout	No	No	Single	Spinous	Spinous and papillate	Capitate	Simple
Amastridium	Round	13–17 + 2	Thin	Yes	Variable	Single, slightly	Spinous	Calyculate	Cap itate ²	Bifurcate, centrolineal
Atractus	Round	5-14	Thin to stout	No	No	Single or bilobed	Spinous	Spinous or calyculate	Noncapitate or	Simple to bifurcate,
Chapinophis	Subcircular	12-13	Thin	No	No	Bilobed	Spinous	Calyculate	Bicapitate	Bifurcate,
Chersodromus	Round	7–9	Very thin to moderate	No	No	Bilobed	Spinous	Spinous	Noncapitate or	centripedal Bifurcate, centrifugal
Coniophanes	Round	8-15 + 2	Thin to moderate	Yes	Yes	Single or bilobed	Spinous	Calyculate or flounced	semicapitate Unicapitate or	Bifurcate, centrolineal
Cryophis	Subcircular ³	19-22 +	Very thin	Yes	No	Single	Spinous	Calyculate	noncapitate Capitate	or centrifugal Bifurcate,
Dipsas articulata	Vertical	$2 \\ 17-24^4$	Very thin	No	No	Slightly	Spinous	Calvculate	Unicapitate	centrolineal Bifurcate,
group Enuliophis	Round	4+2	Very stout	No	No	bilobed Slightly	Spinous	Smooth	Noncapitate	centrolineal Bifurcate
Enulius	Pound	2+2	Vorwatout	No	Variable	bilobed	Spinous	Spinous and	Noncopitate	centrifugal Difurcata
C	Round	3+2	very stout	NO	variable	bilobed	Spinous	papillate	Noncapitate	centrolineal
Geophis omiltemanus group	Round	10-17	Very thin	No	No	bilobed	Spinous	Calyculate	Unicapitate	Bifurcate
<i>Geophis sieboldi</i> group	Round	8–15	Very thin	No	No	Slightly bilobed	Spinous	Calyculate	Unicapitate	Bifurcate
Hydromorphus	Round	11-12 +	Extremely	Yes	No	Bilobed	Spinous	Calyculate and	Unicapitate	Bifurcate,
Hypsiglena (including Eridinhar)	Vertical	7-12 + 2	Very thin to thin	Yes	No	Slightly bilobed	Spinous	Calyculate and spinous	Capitate	Simple
Imantodes	Vertical	10-22 +	Very thin	Yes	Yes	Single	Spinous	Calyculate	Capitate	Simple
Leptodeira	Vertical	8-18 + 2	Very thin to moderate	Variable	Yes	Single	Spinous	Calyculate	Capitate	Simple with tiny terminal fork or bifurcate
Ninia	Round	8-18	Very thin	No	No	Single	Spinous	Calyculate	Capitate	Bifurcate,
Nothopsis	Round	18-19 +	Stout	Yes	No	Single	Spinous and	Spinous	Capitate	Bifurcate,
Omoadiphas	Subcircular	13-14	Very thin	No	No	Bilobed	Spinous	Calyculate	Bicapitate	centrolineal Bifurcate,
Pseudoleptodeira	Vertical	14 + 2	Thin	Yes	No to	Single	Spinous	Spinous	Capitate	centrolineal Simple
latifasciata Rhadinaea	Round	12-22 +	Very thin	Yes	faintly No	Single	Spinous	Calyculate	Capitate	Bifurcate,
decorata group Rhadinaea	Round	2 15–22 +	Thin to	Variable	No	Single	Spinous	Calyculate	Capitate	centrolineal Bifurcate,
<i>taeniata</i> group Rhadinella	Round	2 9-21 + 2 ⁵	moderate Moderate	Variable	No	Bilobed	Spinous	Calyculate	Unicapitate	centrolineal Bifurcate,
Rhadinophanes	Round	19-20 +	Thin	No	No	Bilobed	Spinous	Calyculate	Bicapitate	centrolineal Bifurcate,
Sibon annulatus & S. argus	Vertical	12-17	Thin	No	No	Single	Spinous	Calyculate	Capitate	centripetal Bifurcate, centrolineal
groups Tantalophis	Subcircular	13-17 +	Thin	Yes	No	Bilobed	Spinous	Calyculate	Bicapitate	Bifurcate,
Tretanorhinus	Round	2 27–30	Very thin	No	No	Single	Spinous	Calyculate	Capitate	centripetal Bifurcate,
Trimetopon	Round	11-12 +	Stout	Variable	No	Single or	Usually	Calyculate	Semicapitate	centrolineal Bifurcate,
Tropidodipsas	Su beireu lar	2 12–24	Very thin to	No	No	bilobed Single	spinous Spinous	Calyculate or	Capitate	centrolineal Bifurcate,
Urotheca	Round	11-19+	thin Moderate	Yes	Variable	Single to	Spinous	spinous Calvculate	Unicapitate	centrolineal Bifurcated
(now included within <i>Pliocercus</i>)		2				Slightly bilobed	-1			centrolineal

¹ When "+" inserted between numbers, last number indicating teeth conspicuously enlarged over anterior series; diastema and grooves present or not.

⁴ When "+" inserted between numbers, last number indicating teeth conspicuously enlarged over anterior series; diastema and grooves present or not.
² Report by Wilson and Meyer (1969) of organ being noncapitulate appears in error and at odds with their figure 3.
³ Sensu Myers (1984); distinction between round and subcircular can be slight and further examination of material, especially living, may reveal that some snakes here categorized as having round pupils actually should be judged subcircular. Conversely, some species appearing to have subcircular pupils may contract pupils to the vertical condition under certain circumstances.
⁴ Variation in number of teeth for Middle American species except *Dipsas gaigae*, which is reported to have as few as 10.
⁵ Sometimes more than two posterior teeth enlarged

⁵ Sometimes more than two posterior teeth enlarged.
⁶ Uncertain if *Sibon carri* falls within these groups; this species not examined.

projects upward and forward; the nasal, frontal, and parietal bones have no or little space between them with mostly tight sutures; the quadrate is exceptionally broad, and the maxillary bears 10-11 short, stout teeth. In Geophis anocularis, the premaxilla slopes forward; the nasals, frontals, and parietals are in tight juxtaposition, with no dorsolateral parietal ridge; the frontal forms only a small portion of the orbital socket; no postorbital is present; the quadrate relatively small but expanded dorsally, and the nasals are roughly triangular. In G. laticinctus, the premaxillary is rounded in lateral view; the nasals and frontals are widely separated; the parietal ridge is laterally located; and the maxillary teeth moderately long and stout, decreasing in size posteriorly. In Rhadinaea, the nasals are small, widely separated from the frontals; the postorbital does not contact prefrontals, and the prefrontal is distinctly higher than long. In Sibon and Tropidodipsas, the nasals are widely separated from the frontals, the prefrontal is distinctly higher than long, the postorbital is long and delicate, the dorsolateral ridge of the parietal is well developed, and the quadrate is well developed and robust, with a broadly expanded superior end.

Hemipenes in species of dipsadids often are single or bilobed with a bifurcate sulcus spermaticus, capitate calyculate ornamention on distal portion of lobes, and spines or spinules on hemipenial body and sometimes basal portion of lobes (Fig. 7A,C,D). Genera having a single subcyindrical organ include *Adelphicos, Amastridium, Cryophis, Imantodes, Leptodeira, Ninia, Rhadinaea, Tropidodipsas,* and others (see Table 2), but most of these genera are characterized by a distinctive calyculate apical portion of the hemipenis, which is usually differentiated into a distinct capitulum with a bifurcate sulcus spermaticus. Besides having a single organ, Adelphicos (Fig. 7), *Eridiphas, Hypsiglena, Imantodes, Leptodeira,* and *Pseudoleptodeira* have a simple (not bifurcate) sulcus spermaticus, but these genera differ from *Cenaspis* by having a spinous hemipenial body and some capitation evident.

DISCUSSION

The type locality appears to be very near the Continental Divide, but slightly north on the Atlantic versant. The western flank of Cerro El Baúl is drained by the Río El Baúl and the eastern side by the Río Portamoneda, both upper tributaries of the Río Grijalva flowing into the Gulf of Mexico. A small portion of the southern end of the range is drained by several small streams that are upper tributaries of the Río San Miguel that flows into the Mar Muerto of the Pacific. Cerro El Baúl attains \sim 2,050 m and is the highest point in the mountains of southeastern Oaxaca (although immediately to the east a peak in the Cerro El Ermitaño reaches at least 1,950 m), and is included in the region known at least locally as the Chimalapas, of which Cerro El Baúl and Cerro El Ermitaño can be considered the easternmost outliers. The small settlement and historical collecting locality of Colonia Rodulfo Figueroa, located in the eastern portion of the Chimalapas, has usually been reported to be in Oaxaca (Bogert, 1968; Campbell, 1984). Digital maps provided by Google Earth, however, place Colonia Rodulfo Figueroa in Chiapas ~1 km west of the Oaxaca–Chiapas border.

Although one of the least explored regions in Mexico, the biotic diversity of the Chimalapas is already well recognized (MacDougall, 1971; Smith and Campbell, 1996; Peterson et al., 2003; Aguilar-López et al., 2016), and these highlands contain a high number of herpetofaunal endemics. Moderately high elevations of the Chimalapas, influenced by weather conditions

from both the Gulf of Mexico and Pacific Ocean, have provided the backdrop for a spectacular landscape and biotic inventory. The fortuitous geographical location of the Chimalapas provided the means by which the origin of biota was derived from multiple sources besides the Sierra Madre de Chiapas. These highlands are sometimes considered an extension of the northwestern highlands of the Sierra Madre de Chiapas (Johnson, 1990). Although these highlands lie to the east of the Isthmus of Tehuantepec, a strong biotic component derives from the Mexican Plateau, as well as temperate regions farther north. To the east, the Chimalapas are isolated from the highlands of the Meseta Central of Chiapas and the Sierra Madre del Sur of Chiapas by low ridges and foothills covered with xerophytic vegetation (Leopold, 1950; Miranda, 1952; Breedlove, 1981), but share many highland genera of tropical origin. Perhaps most remarkable are the biotic similarities with the isolated Sierra de Los Tuxtlas (Campbell, 1984), arising out of the Gulf Coast and isolated to the NE by some 150 km of lowland tropical deciduous forest and tropical evergreen forest.

As previously noted, the holotype of *Cenaspis aenigma* was taken from the stomach of a large *Micrurus nigrocinctus* (UTA R-6085). During our exploration of the region, *M. browni* was more common, with about half a dozen individuals encountered; however, only the single *M. nigrocinctus* was found. This Coralsnake was secured by Julio Ornelas-Martínez, a resident of Rizo de Oro who at the time was engaged in harvesting palm (*Chamaedorea*) on the slopes of Cerro Baúl and surrounding uplands.

Although the type of *Cenaspis aenigma* was collected within 20-25 km NE (by road) of Rizo de Oro on the slopes of Cerro Baúl, determining the precise habitat of origin is not possible because of interdigitation of many complex habitats in the area owing to elevation, slope exposure, prevailing winds, and precipitation. The diversity of forest types in this region is described in captivating fashion by MacDougall (1971). Among the various habitats MacDougall (1971) describes is an incredible elfin forest near Cerro Baúl in which trees, shaped by strong and almost constant N-to-S winds, have been beaten down to such an extent that, in places, walking on top of the forest is easier than walking through it. Another unique forest in the region is the cypress woods (Cupressus benthami) covering some of the upper elevations of Cerro Baúl and perhaps elsewhere. Our experience in the region has been that many restricted microhabitats occur in certain ravines and on particular mountain ridges and crests.

Hypothesizing about living *Cenaspis aenigma*, on the basis of morphological traits of the single preserved specimen, is somewhat daunting but worthwhile. The dorsal color is rather unremarkable, being uniformly pale brown. This color and lack of dorsal pattern is not unusual for burrowing species; however, the ventrals are marked with three series of dark rectangular to triangular markings forming essentially three stripes for the length of the body, and the subcaudals are marked with a single midventral band extending the length of the tail. Why a secretive burrowing snake would have such a distinctive ventral pattern is unknown. The ventral pattern is not replicated in any other Middle American snake, although as mentioned previously, several terrestrial (vs. fossorial) species have a vivid pattern of series of spots on the venter.

Snakes are the only terrestrial vertebrates that are usually adapted to swallow large, entire prey often larger than the head of the predator. Because *Cenaspis aenigma* lives in mesic forest, one is tempted to speculate that it preys upon soft-bodied invertebrates such as earthworms and slugs, similar to other sympatric snakes of the genera *Adelphicos, Geophis,* and *Ninia.* Yet all snakes in these genera have long needlelike teeth, in sharp contrast to the short, stout, and blunt teeth of *C. aenigma*. Some of the feeding habits of *C. aenigma* may be implied on the basis of certain features of the skull. The teeth are short, stout, and appear relatively blunt. This type of dentition is characteristic of species that prey on arthropods with hard chitinous exoskeletons and is perhaps most common among the Sonorini (*Sonora, Toluca, Stenorrhina*) that prey on spiders, centipedes, and snakes such as *Symphimus* that feed mainly on arthropods. Relatively few dipsadids possess this dentition, although the teeth in *Rhadinophanes*, some species of *Atractus*, and *Adelphicos* are moderately short but sharp.

Pupil shape may provide some insights into the ecology of snakes (Brischoux et al., 2010). The eyes of the holotype of *Cenaspis aenigma* are in pathetic condition owing to the digestive enzymes of its predator. Nevertheless, they appear subcircular in shape. Brischoux et al. (2010) suggest that round-pupil species adapted to low light conditions are unable to close the pupil sufficiently to prevent dazzle under daylight conditions, in contrast with vertical-pupil species that may have better vision both at night and by day. It follows that visual acuity for small-eyed species with either round or subcircular pupils is greatly diminished during daylight hours and these species are highly secretive, remaining underground almost without interruption or perhaps occasionally venturing out by night, especially after rains.

The overall morphology of C. aenigma is that of a burrowing snake. Cenaspis aenigma is similar to the burrowing genera Geophis, Adelphicos, and Atractus in having an elongate braincase. Categorizing relative length of the snake skull into long vs. short braincase is a somewhat subjective assessment, but cursorial examination does reveal conspicuous differences in relative head length/width that probably associates with evolutionary histories and ecologies. We used only adult snakes in our examinations because of the large amount of variation attributable to allometry (Murta-Fonseca and Fernandes, 2016). If the distance from the level of the parietal-frontal suture above the eye to the posterior edge of the exoccipital midline suture is divided by the distance across the parietal at its widest point in dorsal aspect (usually about midlevel), then fossorial species with elongate braincases produce values of 0.48-0.60 (Cenaspis is 0.59, Adelphicos 0.54-0.56, Atractus 0.48-0.59); those for terrestrial species vary from 0.65 to 0.80 (Imantodes 0.80, Leptodeira 0.65, Ninia 0.67, Tropidodipas 0.66, Rhadinaea 0.67-0.71). The genus Geophis contains some groups that are highly fossorial and others that are more terrestrial. Species of this genus may fall into either long or short braincase categories, using 0.63 as the arbitrary dividing line.

Characteristics of "long-headed burrowers" often include long postorbital braincases, shortened orbits, low prefrontals, short or missing postorbitals, reduced postorbital processes on parietal, anterior extensions of the dorsolateral edge of parietal along lateral edge of frontal, elongated parietal, shortening of caudal end of the supratemporal, and reductions of transversely oriented crests associated with muscle origins (Cundall and Irish, 2008). Most of these features are possessed by *Cenaspis aenigma*, but the postorbitals are relatively well developed and the dorsolateral ridge is only moderately well developed, merging with a prominent ridge forming part of the dorsal articulation with the postorbital. Given its unique features, the affinities of *Cenaspis aenigma* are difficult to assess. The only other genus of New World colubrid with undivided subcaudals is *Rhinocheilus*, but invariably at least a few subcaudals (usually distally) are divided. That genus is a member of the Colubridae and the differences between it and *Cenaspis* are numerous and striking. Very superficially, *Cenaspis* seems to be allied with *Adelphicos* or *Geophis*, widespread groups showing great morphological and ecological diversity. Yet, osteological features, the enigmatic entire subcaudals, and unique hemipenis of *Cenaspis* preclude its placement in these genera and suggest that the relationships of *Cenaspis* are distant and will have to be found elsewhere.

Acknowledgments.—We are grateful to the late J. Ornelas-Martínez, who collected the type of the snake described herein and who saved one of us (JAC) from unpleasant situations on several occasions, including once when our vehicle had fallen into a 15-foot-deep ravine as storm clouds gathered to the north. We thank the Comisión Nacional para el Conocimiento de la Biodiversidad for permission to conduct studies in Mexico. Several National Science Foundation grants (DEB-0613802 and 0102383 to JAC; DEB-0416160 to ENS) allowed us to conduct fieldwork on Mexico's incredible herpetodiversity, but we were unfortunately unsuccessful in obtaining additional material of *Cenaspis* despite repeated attempts. We thank M. Loocke and the Shimadzu Institute for allowing complimentary use of the SMX-100CT scanner. A Beta Phi chapter Phi Sigma research grant to ASH funded the CT reconstruction computer used for this work.

LITERATURE CITED

- AGUILAR-LÓPEZ, J. L., E. PINEDA, R. LUÍA-MANZANO, AND L. CANSECO-MÁRQUEZ. 2016. Species diversity, distribution, and conservation status in a Mesoamerican region: amphibians of the Uxpanapa-Chimalapas Region, Mexico. Tropical Conservation Science 2016:1– 16.
- ANGARITA-SIERRA, T. 2014. Hemipenial morphology in the semifossorial snakes of the genus *Ninia* and a new species from Trinidad, West Indies (Serpentes: Dipsadidae). South American Journal of Herpetology 9:114–130.
- BOGERT, C. M. 1968. A new arboreal pit viper of the genus *Bothrops* from the Isthmus of Tehuantepec, Mexico. American Museum Novitates 2341:1–14.
- BOGERT, C. M., AND W. E. DUELLMAN. 1963. A new genus and species of colubrid snake from the Mexican state of Oaxaca. American Museum Novitates 2162:1–15.
- BREEDLOVE, D. L. 1981. Flora of Chiapas. Part 1. Introduction to the Flora of Chiapas. California Academy of Sciences, USA.
- BRISCHOUX, F., L. PIZZATTO, AND R. SHINE. 2010. Insights into the adaptive significance of vertical pupil shape in snakes. Journal of Evolutionary Biology 23:1878–1885.
- BURGER, W. L., AND J. E. WERLER. 1954. The subspecies of the ring-necked coffee snake, *Ninia diademata*, and a short biological and taxonomic account of the genus. University of Kansas Science Bulletin 36:643– 672.
- CADLE, J. E. 1984a. Molecular systematics of Neotropical Xenodontine snakes: I. South American Xenodontines. Herpetologica 40:8–20.
- CADLE, J. E. 1984b. Molecular systematics of Neotropical Xenodontine snakes: II. Central American Xenodontines. Herpetologica 40:21–30.
- CADLE, J. E. 1984c. Molecular systematics of Neotropical Xenodontine snakes: III. Overview of Xenodontine phylogeny and history of New World snakes. Copeia 1984:641–652.
- CAMPBELL, J. A. 1984. A new species of *Abronia* (Sauria: Anguidae) with comments on the herpetogeography of the highlands of southern Mexico. Herpetologica 40:373–381.
- CAMPBELL, J. A. 1989. A new species of colubrid snake of the genus *Coniophanes* from the highlands of Chiapas, Mexico. Proceedings of the Biological Society of Washington 102:1036–1044.

- CAMPBELL, J. A., AND E. D. BRODIE, JR. 1988. A new colubrid snake of the genus Adelphicos from Guatemala. Herpetologica 44:416–422.
- CAMPBELL, J. A., AND L. S. FORD. 1982. Phylogenetic relationships of the colubrid snakes of the genus *Adelphicos* in the highlands of Middle America. Occasional Papers of the Museum of Natural History University of Kansas 100:1–22.
- CAMPBELL, J. A., AND E. S. SMITH. 1998. A new genus and species of colubrid snake from the Sierra de las Minas of Guatemala. Herpetologica 54:207–220.
- CANSECO-MÁRQUEZ, L., C. G. RAMÍREZ-GONZÁLEZ, AND J. A. CAMPBELL. 2018. Taxonomic review of the rare Mexican snake genus *Chersodromus* (Serpentes: Dipsadidae), with the description of two new species. Zootaxa 4399:151–169.
- COPE, E. D. 1894. The classification of snakes. American Naturalist 28: 831–844.
- CROTHER, B. I. 1989. A redescription of the hemipenis of *Hydromorphus* concolor (Colubridae) with comments on its tribal allocation. Copeia 1989:227–229.
- CUNDALL, D. 1981. Cranial osteology of the colubrid snake *Opheodrys*. Copeia 1981:353–371.
- CUNDALL, D., AND F. IRISH. 2008. The snake skull. Pp. 349–692 in C. Gans (ed.), Biology of the Reptilia. Volume 20, Morphology H, The Skull of the Lepidosauria. Society for the Study of Amphibians and Reptiles, USA.
- DOWLING, H. G. 1951. A proposed standard system for counting ventrals in snakes. British Journal of Herpetology 1:97–99.
- DowLING, H. G. 1967. Hemipenes and other characters in colubrid classification. Herpetologica 23:138–142.
- DOWLING, H. G., AND J. M. SAVAGE. 1960. A guide to the snake hemipenis: a survey of basic structure and systematic characteristics. Zoologica 45:17–27.
- DOWNS, F. L. 1967. Intrageneric relationships among colubrid snakes of the genus *Geophis* Wagler. Miscellaneous Publications of the Museum of Zoology University of Michigan 131:1–193.
- DUELLMAN, W. E. 1958a. Systematic status of the colubrid snake, Leptodeira discolor Günther. University of Kansas Publications Museum of Natural History 11:1–9.
- DUELLMAN, W. E. 1958b. A monographic study of the colubrid snake genus *Leptodeira*. Bulletin of the American Museum of Natural History 114:1–151.
- FIGUEROA, A., A. D. MCKELVY, L. L. GRISMER, C. D. BELL, AND S. P. LAILVAUX. 2016. A species-level phylogeny of extant snakes with description of a new colubrid subfamily and genus. PLoS ONE 11(9): e0161070.
- GOOGLE EARTH PRO. 2017. Map Data: Digital Globe. Imagery date 15 April 2009. Available at https://www.google.com/earth/desktop/.
- GRAZZIOTIN, F. G., H. ZAHER, R. W. MURPHY, G. SCROCCHI, M. A. BENAVIDES, Y. ZHANG, AND S. L. BONATTO. 2012. Molecular phylogeny of the New World Dipsadidae (Serpentes, Colubroidea): a reappraisal. Cladistics 28:437–459.
- HARVEY, M. B., G. RIVAS-FUENMAYOR, J. R. CAICEDO, AND J. V. RUEDA-ALMONACID. 2008. Systematics of the enigmatic dipsadine snake *Tropidodipsas perijanensis* Alemán (Serpentes: Colubridae) and review of morphological characters of Dipsadini. Herpetological Monographs 22:106–132.
- HAY, O. P. 1892. Description of a supposed new species of *Storeria* from Florida, *Storeria victa*. Science 19:199.
- INSTITUTO NACIONAL DE ESTADÍSTICA, GEOGRAFÍA E INFORMÁTICA, MÉXICO. 2000. Carta Topográfica 1:250,000, Salina Cruz E15–10, D15–1, segunda impresión.
- JENNER, J. V., AND H. G. DOWLING. 1985. Taxonomy of American xenodontine snakes: the tribe Pseudoboini. Herpetologica 41:161– 172.
- JOHNSON, J. D. 1990. Biogeographic aspects of the herpetofauna of the Central Depression of Chiapas, México, with comments on surrounding areas. Southwestern Naturalist 35:268–278.
- KLACZKO, J., E. SHERRAT, AND E. Z. F. SETZ. 2016. Are diet preferences associated to skulls shape diversification in xenodontine snakes? PLoS ONE 11:1–12.
- KOFRON, C. P. 1985a. Review of the Central American colubrid snakes, Sibon fischeri and S. carri. Copeia 1985:164–174.
- KOFRON, C. P. 1985b. Systematics of the Neotropical gastropod-eating snake genera, *Tropidodipsas* and *Sibon*. Journal of Herpetology 19:84– 92.
- KOFRON, C. P. 1987. Systematics of Neotropical gastropod-eating snakes: the *faciata* group of the genus *Sibon*. Journal of Herpetology 21:210– 225.

LEOPOLD, A. S. 1950. Vegetation zones of Mexico. Ecology 31:507–518.

- LEVITON, A. E., AND W. W. TANNER. 1960. The generic allocation of *Hypsiglena slevini* Tanner (Serpentes: Colubridae). Occasional Papers of the California Academy of Sciences 27:1–7.
- MacDougall, T. 1971. The Chima wilderness. Explorers' Journal 1971: 86–103.
- McDowell, S. B. 2008. The skull of serpents. Pp. 467–620 in A. S. Gaunt and K. Adler (eds.), Biology of the Reptilia. Volume 21, Morphology I, The Skull and Appendicular Locomotor Apparatus of Lepidosauria. Society for the Study of Amphibians and Reptiles, USA.
- MIRANDA, F. 1952. La Vegetación de Chiapas, 1. Ediciones Gobierno de Estado. Chiapas, Mexico.
- MULCAHY, D. G. 2003. Geographic variation in the Baja California nightsnake (*Eridiphas slevini*), with comments on taxonomy and diet. Journal of Herpetology 37:566–571.
- MULCAHY, D. G. 2007. Molecular systematics of Neotropical cat-eyed snakes: a test of the monophyly of Leptodeirini (Colubridae: Dipsadinae) with implication for character evolution and biogeography. Biological Journal of the Linnean Society 92:483–500.
- MULCAHY, D. G., T. H. BECKSTEAD, AND J. SITES. 2011. Molecular systematics of the Leptodeirini (Colubroidea: Dipsadidae) revisited: species tree analyses and multi-locus data. Copeia 2011:407–417.
- MURTA-FONSECA, R. A., AND D. S. FERNANDES. 2016. The skull of *Hydrodynastes gigas* (Duméril, Bibron, and Duméril, 1854) (Serpentes: Dipsadidae) as a model of snake ontogenetic allometry inferred by geometric morphometrics. Zoomorphology 135:233–241.
- MYERS, C. W. 1974. The systematics of *Rhadinaea* (Colubridae), a genus of New World snakes. Bulletin of the American Museum of Natural History 153:1–262.
- MYERS, C. W. 1982. Blunt-headed vine snakes (*Imantodes*) in Panama, including a new species and other revisionary notes. American Museum Novitates 2738:1–50.
- MYERS, C. W. 1984. Subcircular pupil shape in the snake *Tantaophis* (Colubridae). Copeia 1984:215–216.
- MYERS, C. W. 2003. Rare snakes—five new species from eastern Panama: reviews of northern *Atractus* and southern *Geophis* (Colubridae: Dipsadinae). American Museum Novitates 3391:1–47.
- MYERS, C. W., AND J. E. CADLE. 2003. On the snake hemipenis, with notes on *Psomophis* and techniques of eversion: a response to Dowling. Herpetological Review 34:295–302.
- MYERS, C. W., AND J. A. CAMPBELL. 1981. A new genus and species of colubrid snake from the Sierra Madre del Sur of Guerrero, Mexico. American Museum Novitates 2708:1–20.
- NIETO MONTES DE OCA, A. 2003. A new species of the *Geophis dubius* group (Squamata: Colubridae) from the Sierra de Juárez of Oaxaca, Mexico. Herpetologica 59:572–585.
- PASSOS, P., J. C. ARREDONTO, R. FERNANDES, AND J. D. LYNCH. 2009. Three new *Atractus* (Serpentes: Dipsadidae) from the Andes of Colombia. Copeia 2009:425–436.
- PASSOS, P., M. T. JUNIOR, R. S. RECODER, M. A. DE SENA, F. D. VECHIO, H. B. DE A. PINTO, S. H. S. T. MENDOÇA, J. CASSIMIRO, AND M. T. RODRIGUES. 2013. A new species of *Atractus* (Serpentes: Dipsadidae) from Serra do Cipó, Espinhaço Range, Southeastern Brazil, with proposition of a new species group to the genus. Papéis Avulsos de Zoologia 53:75– 85.
- PESANTES, O. S. 1994. A method for preparing the hemipenis of preserved snakes. Journal of Herpetology 28:93–95.
- PETERS, J. A. 1960. Dictionary of Herpetology. Hafner Publishing Company, USA.
- PETERS, J. A. 1964. Supplemental notes on snakes of the subfamily Dipsadinae (Reptilia: Colubriadae). Studies on Neotropical Fauna and Environment 4:45–50.
- PETERSON, A.T, NAVARRO-SIGÜENZA, A.G., HERNÁNDEZ-BAÑOS, B.E., ESCALO-NA-SEGURA, G., RABÓN-GALLARDO, F., RODRÍGUEZ-AYALA, E., FIGUEROA-ESQUIVEL, E.M., AND CABRERA-GARCÍA, L. 2003. The Chimalapas Region, Oaxaca, Mexico: a high-priority region for bird conservation in Mesoamerica. BirdLife International 127:227–253.
- PIETRO, D. O. D., L. ALCALDE, AND J. D. WILLIAMS. 2014. New cranial characters in the tribe Hydropsini (Serpentes: Dipsadidae: Xenodontinae). Acta Herpetologica 9:1–14.
- PINOU, T., S. VICARIO, M. MARSCHNER, AND A. CACCONE. 2004. Relict snakes of North America and their relationships within Caenophidia, using likelihood-based Bayesian methods on mitochondrial sequences. Molecular Phylogenetics and Evolution 32:563–574.
- SAVAGE, J. M. 2002. The Amphibians and Reptiles of Costa Rica. The University of Chicago Press, USA.

- SCHARGEL, W. E., AND T. A. CASTOE. 2003. The hemipenes of some snakes of the semifossorial genus *Atractus*, with comments on variation in the genus. Journal of Herpetology 37:718–721.
- SCHMIDT, K. P. 1932. Stomach contents of some American coral snakes, with the description of a new species of *Geophis*. Copeia 1932:6–9.
- SMITH, B. E., AND J. A. CAMPBELL. 1996. The systematic status of Guatemalan populations of snakes allied with *Ninia maculata* (Reptilia: Colubridae). Proceedings of the Biological Society of Washington 109:749–754.
- SMITH, H. M. 1941. On the Mexican snakes of the genus *Pliocercus*. Proceedings of the Biological Society of Washington 54:119–124.
- SMITH, H. M., AND D. CHISZAR. 2001. Reassessment of the nominal species of False Coral Snakes, *Pliocercus psychoides* (Reptilia: Serpentes). Southwestern Naturalist 46:114–116.
- SMITH, H. M., AND E. H. TAYLOR. 1945. An annotated checklist and key to the snakes of Mexico. Bulletin of the United States National Museum 187:1–239.
- TANNER, W. W. 1943. Two new species of *Hypsiglena* from western North America. Great Basin Naturalist 4:49–55.
- TANNER, W. W. 1944. A taxonomic study of the genus *Hypsiglena*. Great Basin Naturalist 5:25–90.
- TOWNSEND, J. H. 2006. *Geophis dunni* Schmidt. Catalogue of American Amphibians and Reptiles 838:1–3.
- VIDAL, N., A. S. DELMAS, P. DAVID, C. CRUAUD, A. COULOUX, AND S. B. HEDGES. 2007. The phylogeny and classification of caenophidian snakes inferred from seven nuclear protein-coding genes. Comtes Rendus Biologies 330:182–187.
- VIDAL, N., M. DEWYNTER, AND D. J. GOWER. 2010. Dissecting the major American snake radiation: a molecular phylogeny of the Dipsadidae

Bonaparte (Serpentes: Caenophidia). Comptes Rendus Biologies 333: 48–55.

- WILSON, L. D., AND J. R. MEYER. 1969. A review of the colubrid snake genus *Amastridium*. Bulletin of the Southern California Academy of Sciences 68:146–160.
- ZAHER, H. 1999. Hemipenial morphology of the South American xenodontine snakes, with a proposal for a monophyletic Xenodontinae and a reappraisal of colubroid hemipenes. Bulletin of the American Museum of Natural History 240:1–168.
- ZAHER, H., AND A. L. C. PRUDENTE. 2003. Hemipenes of Syphlophis (Serpentes, Xenodontidae) and techniques of hemipenial preparation in snakes: a response to Dowling. Herpetological Review 34:302–307.
- ZAHER, H., F. G. GRAZZIOTIN, J. E. CADLE, J. E. MURPHY, J. C. MOURA-LEITE, AND J. C. BONATTO. 2009. Molecular phylogeny of advanced snakes (Serpentes, Caenophidia) with an emphasis on South American Xenonodontines: a revised classification and descriptions of new taxa. Papéis Avulsos de Zoologia 49:115–153.
- ZAHER, H., L. DE OLIVEIRA, F. G. GRAZZIOTIN, M. CAMPAGNER, C. JARED, M. M. ANTONIAZZI, AND A. L. PRUDENTE. 2014. Consuming viscous prey: a novel protein-secreting delivery system in Neotropical snail-eating snakes. BMC Evolutionary Biology 14:58.

Accepted: 17 September 2018. Published online: 27 November 2018.

ZooBank ID: lsid:zoobank.org:pub:981A3DDB-53F9-49D2-B3CD-874A1CEF84C0